

## BCMB 3100 – Chapter 3 (part 1)

- Diversity of protein function
- Complete definition of amino acids
- Memorize complete structure of 20 common amino acids!!!
- pKa's of  $\alpha$  amino and  $\alpha$  carboxyl groups
- Amino acids with ionizable side groups
- Titration curve

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## PROTEIN STRUCTURE AND FUNCTION

**Mulder** (1800s): showed albumins (milk and egg) contained \_\_\_\_\_

**Berzelius** (1838): named such substances

\_\_\_\_\_ *from Greek proteios "of first rank"*

**Proteins** recognize and \_\_\_\_\_ many different types of molecules & \_\_\_\_\_ most of the \_\_\_\_\_ necessary for life.

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## Examples of Protein Function

\_\_\_\_\_ :  
enzymes increase reaction rates by  $\geq 10^6$ -fold; nearly all known enzymes are proteins

\_\_\_\_\_ :  
many small molecules and ions are transported by proteins;  
examples: hemoglobin, myoglobin

\_\_\_\_\_ :  
examples: myosin and actin (muscle movement)

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## Examples of Protein Function

\_\_\_\_\_ : collagen (tensile strength to skin and bones)

\_\_\_\_\_ : antibodies

\_\_\_\_\_ : example: acetylcholine receptor

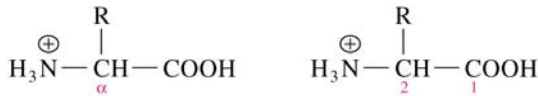
\_\_\_\_\_ :  
DNA binding proteins (repressors, activators, transcription factors, hormones, regulation of translation)

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### WHAT ARE PROTEINS?

Proteins are macromolecules made up of amino acids (\_\_\_\_\_)

$\alpha$ -amino acids: consist of an \_\_\_\_\_, a \_\_\_\_\_, a \_\_\_\_\_ and a distinctive \_\_\_\_\_ bonded to a carbon atom. This carbon is called the \_\_\_\_\_ because it is adjacent to the carboxyl group.



### BCMB 3100 - Chapter 3

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- Titration curves & pI

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Amino acids in solution will be in a charged state. The amino group and/or the carboxyl group will be charged depending upon the pH. The R group may also be charged.

At neutral pH amino acids are predominantly \_\_\_\_\_ (zwitterions)

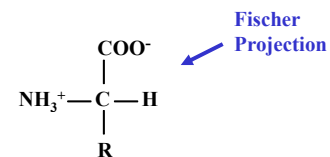
- Under normal cellular conditions amino acids are **zwitterions** (dipolar ions):

Amino group =  **$-\text{NH}_3^+$**

Carboxyl group =  **$-\text{COO}^-$**

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### General Structure of the ionized form of an amino acid



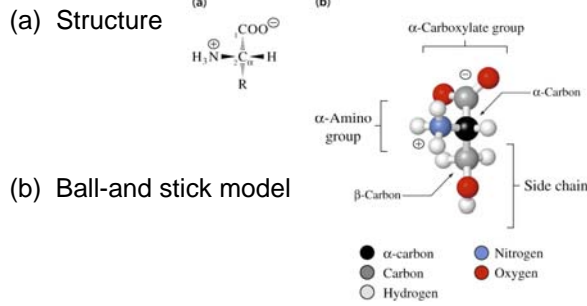
$\alpha$  COO<sup>-</sup>: pKa 1.8-2.5

$\alpha$  NH<sup>+</sup>: pKa 8.7-10.7

**You MUST know this!!!**

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**Fig 3.1 Two representations of an amino acid at neutral pH**



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Amino acids are \_\_\_\_\_ (asymmetric) due to the tetrahedral array of 4 different groups around the  $\alpha$ -carbon (glycine is an exception).

Thus all amino acids except glycine can exist as enantiomers: two stereoisomers that are nonsuperimposable mirror images of each other.

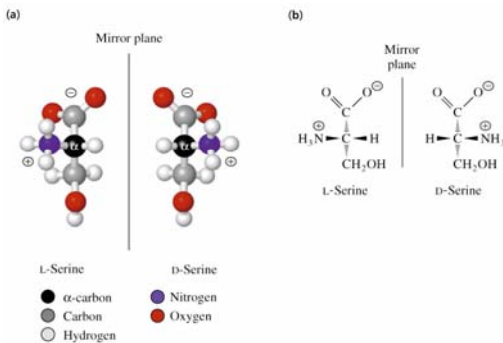
Enantiomers of amino acids are called D (right-handed) or L (left-handed)

L and D refer to absolute configuration

\_\_\_\_\_ are the only constituents of proteins

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**Fig 3.2 Mirror-image pairs of amino acids**



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20 different amino acids are found in proteins

\_\_\_\_\_ (side chains) that differ in size, shape, charge, hydrogen-bonding capacity & chemical reactivity → 20 different amino acids found in proteins of all organisms from bacteria to humans

The amino acid alphabet is at least 2 billion years old

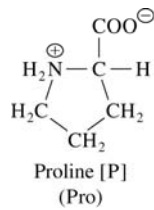
The diversity of protein structure & function is due to the sequence and number of amino acids found in a protein (primary structure)

It is essential that a biochemist commit to memory the structure of the 20 amino acids!  
Know this!! (structure, & 1 & 3 letter code)



## Proline has a nitrogen in the \_\_\_\_\_ ring system

- **Proline (Pro, P)** - has a three carbon side chain bonded to the  $\alpha$ -amino nitrogen
- The heterocyclic pyrrolidine ring restricts the geometry of polypeptides



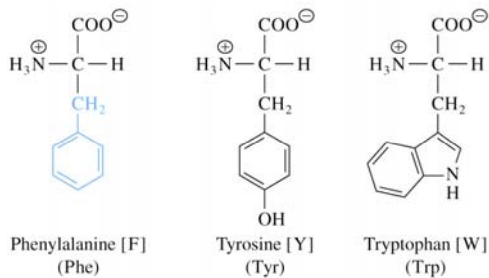
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## \_\_\_\_\_ R Groups

- Side chains have aromatic groups
- **Phenylalanine (Phe, F)** - benzene ring (OD260 nm)
- **Tyrosine (Tyr, Y)** - phenol ring (OD280 nm)
- **Tryptophan (Trp, W)** - bicyclic indole group (OD280 nm)

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## Aromatic amino acid structures



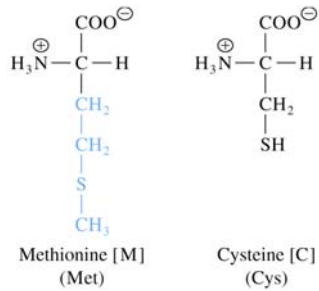
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## \_\_\_\_\_ R Groups

- **Methionine (Met, M)** - (-CH<sub>2</sub>CH<sub>2</sub>SCH<sub>3</sub>)
- **Cysteine (Cys, C)** - (-CH<sub>2</sub>SH)
- Two cysteine side chains can be cross-linked by forming a disulfide bridge (-CH<sub>2</sub>-S-S-CH<sub>2</sub>-)
- Disulfide bridges may stabilize the three-dimensional structures of proteins

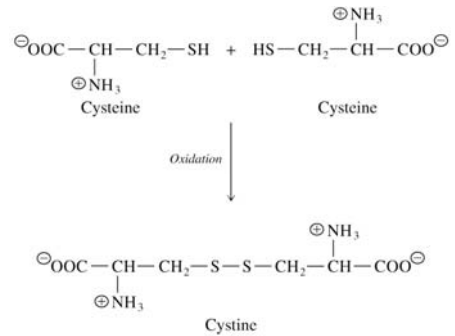
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## Methionine and cysteine



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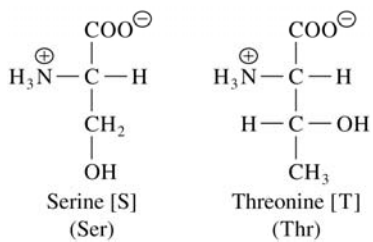
Fig 3.4 Formation of cystine



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## Side Chains with \_\_\_\_\_

**Serine (Ser, S)** and **Threonine (Thr, T)** have uncharged polar side chains



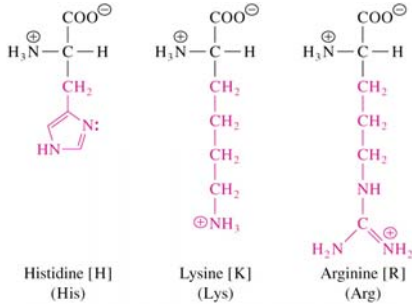
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## \_\_\_\_\_ R Groups

- **Histidine (His, H)** - imidazole
- **Lysine (Lys, K)** - alkylamino group
- **Arginine (Arg, R)** - guanidino group
- Side chains are nitrogenous bases which are substantially positively charged at pH 7 (true for K & R) [ NOTE: error in book, pg 59 ]

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### Structures of histidine, lysine and arginine



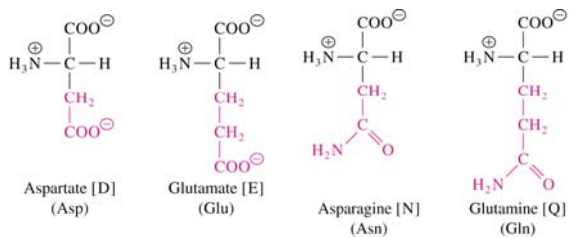
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### \_\_\_\_\_ R Groups and \_\_\_\_\_ Derivatives

- **Aspartate (Asp, D)** and **Glutamate (Glu, E)** are dicarboxylic acids, and are negatively charged at pH 7
- **Asparagine (Asn, N)** and **Glutamine (Gln, Q)** are uncharged but highly polar

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### Structures of aspartate, glutamate, asparagine and glutamine



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### \_\_\_\_\_ of Amino Acid Side Chains

- \_\_\_\_\_: the relative hydrophobicity of each amino acid
- The larger the hydrophathy, the greater the tendency of an amino acid to prefer a hydrophobic environment
- Hydrophathy affects protein folding:  
 \_\_\_\_\_ chains tend to be in the \_\_\_\_\_  
 \_\_\_\_\_ tend to be on the \_\_\_\_\_

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## Challenge of the Week

given out on 8-20-08

Work with your group of 4 people and find at least one example of a mutation in humans, or in industry-relevant plants, animals or microbes. Present, on a single, one-side, typed page, the amino acid mutated, the phenotype of the effect on the organism, the molecular reason that the mutation causes the effect(s), and the effect that this mutation has on/for humans.

Hand in a single, one-sided, typed sheet of paper with ALL group members names (first and last name correctly spelled) as well as a single sentence behind each name describing their contribution to the answer. **These will be collected ONLY at the Breakout Session on August 31.**

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## \_\_\_\_\_ of Amino Acid Side Chains

- \_\_\_\_\_: the relative hydrophobicity of each amino acid
- The larger the hydrophathy, the greater the tendency of an amino acid to prefer a hydrophobic environment
- Hydrophathy affects protein folding:
  - \* \_\_\_\_\_ side chains tend to be in the interior
  - \* \_\_\_\_\_ residues tend to be on the surface

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Table 3.1

- Hydrophathy scale for amino acid residues

(Free-energy change for transfer of an amino acid from interior of a lipid bilayer to water)

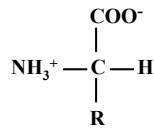
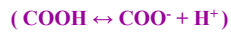
Amino acid	Free-energy change for transfer (kJmol <sup>-1</sup> )
<u>Highly hydrophobic</u>	
Isoleucine	3.1
Phenylalanine	2.5
Valine	2.3
Leucine	2.2
Methionine	1.1
<u>Less hydrophobic</u>	
Tryptophan	1.5*
Alanine	1.0
Glycine	0.67
Cysteine	0.17
Tyrosine	0.08
Proline	-0.29
Threonine	-0.75
Serine	-1.1
<u>Highly hydrophilic</u>	
Histidine	-1.7
Glutamate	-2.6
Asparagine	-2.7
Glutamine	-2.9
Aspartate	-3.0
Lysine	-4.6 <sup>31</sup>
Arginine	-7.5

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Amino acids in solution will be in a \_\_\_\_\_. The amino group and/or the carboxyl group will be charged depending upon the pH. The R group may also be charged. At neutral pH amino acids are predominantly dipolar ions (\_\_\_\_\_).



Note:  
pH dependence  
of the ionization  
state

$\alpha$  COO<sup>-</sup>: pKa 1.8-2.5

$\alpha$  NH<sup>+</sup>: pKa 8.7-10.7

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## Ionization of Amino Acids

- Ionizable groups in amino acids: (1)  $\alpha$ -carboxyl, (2)  $\alpha$ -amino, (3) some side chains
- Each ionizable group has a specific pK<sub>a</sub>



- For a solution pH below the pK<sub>a</sub>, the protonated form predominates (**AH**)
- For a solution pH above the pK<sub>a</sub>, the unprotonated (conjugate base) form predominates (**B**)

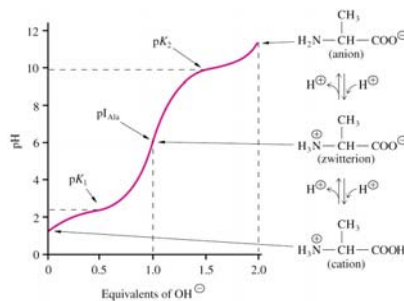
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Fig 3.6 Titration curve for alanine

- Titration curves are used to determine pK<sub>a</sub> values
- pK<sub>1</sub> = 2.4
- pK<sub>2</sub> = 9.9
- pI<sub>Ala</sub> = isoelectric point

(pI = pH when net charge is zero)

pI = \_\_\_\_\_



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## pK<sub>a</sub> values of amino acids

All amino acids contain two ionizable groups:  
 $\alpha$ -carboxyl (pKa 1.8-2.5)  
and protonated  $\alpha$ -amino group (pKa 8.7-10.7)  
(Table 3.2, pg 64 gives specific pKa values)

The \_\_\_\_\_ are also ionizable.  
The pK<sub>a</sub> of these R groups can range from 3.9 to 12.5. You must memorize the pK<sub>a</sub>'s of the side chains of these seven amino acids!

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Table 3.2

Amino acid	pKa value		
	$\alpha$ Carboxyl group	$\alpha$ -Amino group	Side chain
Glycine	2.4	9.8	
Alanine	2.4	9.9	
Valine	2.3	9.7	
Leucine	2.3	9.7	
Isoleucine	2.3	9.8	
Methionine	2.1	9.3	
Proline	2.0	10.6	
Phenylalanine	2.2	9.3	
Tryptophan	2.5	9.4	
Serine	2.2	9.2	
Threonine	2.1	9.1	
Cysteine	1.9	10.7	8.4
Tyrosine	2.2	9.2	10.5
Asparagine	2.1	8.7	
Glutamine	2.2	9.1	
Aspartic acid	2.0	9.9	3.9
Glutamic acid	2.1	9.5	4.1
Lysine	2.2	9.1	10.5
Arginine	1.8	9.0	12.5
Histidine	1.8	9.3	6.0

pK<sub>a</sub> values of amino acid ionizable groups

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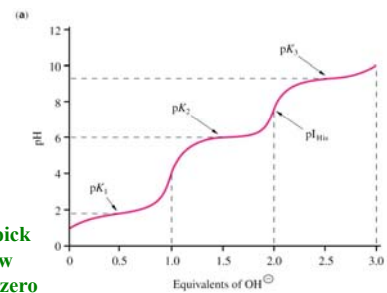
Fig 3.7 (pg 64) Ionization of Histidine

(a) Titration curve of histidine

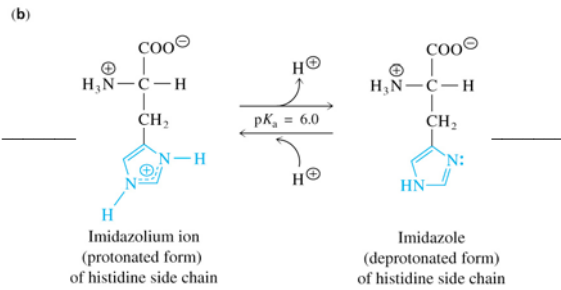
$$\begin{aligned} pK_1 &= 1.8 \\ pK_2 &= 6.0 \\ pK_3 &= 9.3 \end{aligned}$$

pI = pH at which net charge is zero

To determine the pI: pick the pKa above & below point where charge is zero and average those 2 pKa's



**Fig 3.7 (b) Deprotonation of imidazolium ring**



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**Fig 3.7 (pg 64) Ionization of Histidine**

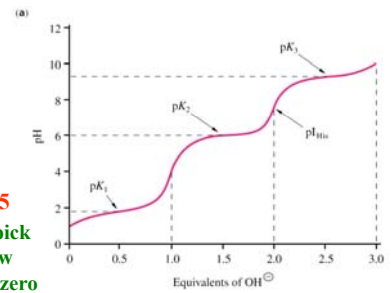
(a) Titration curve of histidine

$pK_1 = 1.8$   
 $pK_2 = 6.0$   
 $pK_3 = 9.3$

$pI = pH$  at which net charge is zero

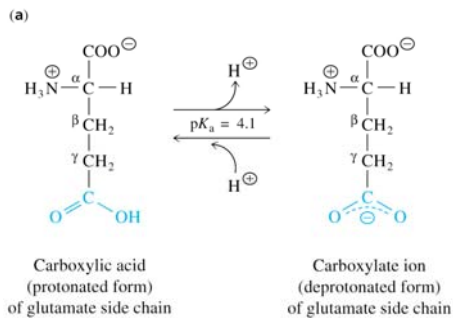
$pI = (6 + 9.3)/2 = 7.65$

To determine the pI: pick the pKa above & below point where charge is zero and average those 2 pKa's



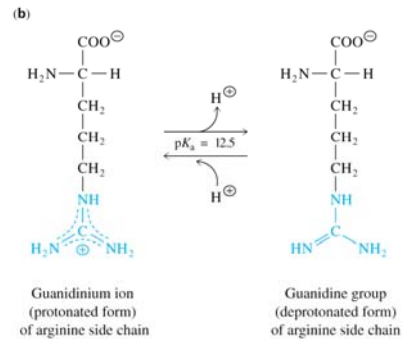
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**Fig 3.8 (a) Ionization of the protonated  $\gamma$ -carboxyl of glutamate**



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**Fig 3.8 (b) Deprotonation of the guanidinium group of Arg**



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## BCMB 3100 – Chapter 3 (part 2)

- Summary of amino acids
- Polypeptides: definition, structure, and direction
- Peptide bond
- Disulfide bonds
- Protein purification
- Methods to determine amino acid composition, cleavage of proteins, protein sequencing
- Diversity of proteins

## The 20 amino acid are divided into 7 groups

- Aliphatic: Gly, Ala, Val, Leu, Ile, Pro
- Aromatic: Phe, Tyr, Trp
- Sulfur-containing: Met, His
- Alcohols: Ser, Thr
- Basic: His, Lys, Arg
- Acidic: Asp, Glu
- Amides: Asn, Gln

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## BCMB 3100 - Chapter 3

- Quick review on amino acids
- Polypeptides: definition, structure, and direction
- Disulfide bonds
- Protein purification
- Methods to determine amino acid composition, cleavage of proteins, protein sequencing

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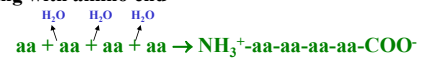
Amino acids are linked by peptide bonds to form polypeptide chains

$\alpha$  carboxyl of one amino acid is joined to  $\alpha$ -amino group of another amino acid by a peptide bond (amide bond) with concomitant loss of  $H_2O$

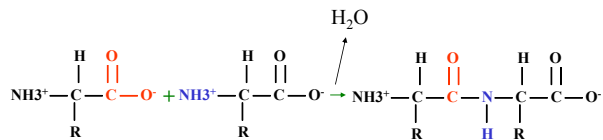
Many amino acids are joined by peptide bonds to form an unbranched polypeptide

Residue: amino acid unit in a polypeptide

By convention the direction of a polypeptide is written starting with amino end



## Formation of a dipeptide



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Most polypeptides contain between \_\_\_\_\_ amino acids

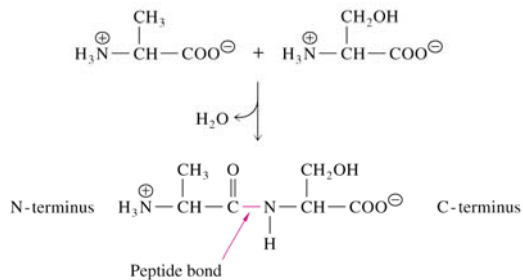
Average M.W. for an amino acid is \_\_\_\_\_ so M.W. of most proteins is 5500 to 220,000 daltons. (One dalton equals one atomic mass unit; kilodalton = 1000 daltons). Most proteins have M.W. of 5.5-220 kd.

Some proteins contain disulfide bonds that cross-link between \_\_\_\_\_ residues by the oxidation of \_\_\_\_\_. The resulting disulfide is called cystine.

Intracellular proteins often lack disulfides while extracellular proteins often have them.

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**Fig 3.9** Peptide bond between two amino acids

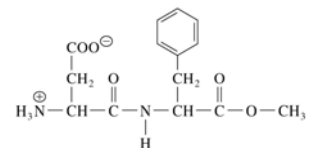


Formation of peptide bonds eliminates the ionizable  $\alpha$ -carboxyl and  $\alpha$ -amino groups of the free amino acids except for those at the amino and carboxy termini

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**Fig 3.10** Aspartame, an artificial sweetener

- Aspartame is a dipeptide methyl ester (aspartylphenylalanine methyl ester)
- About 200 times sweeter than table sugar
- Used in diet drinks



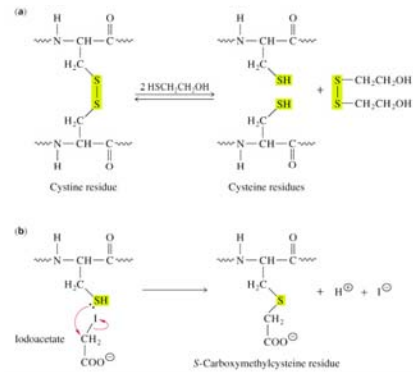
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### BCMB 3100 - Chapter 3

- Memorize complete structure of 20 common amino acids!!!
- pKa's of  $\alpha$  amino and  $\alpha$  carboxyl groups
- Amino acids with ionizable side groups
- Titration curves & pI
- Polypeptides: definition, structure, and direction
- Peptide bond
- **Disulfide bonds**
- Protein purification
- Methods to determine amino acid composition, cleavage of proteins, protein sequencing

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**Fig 3.19** Cleaving, blocking disulfide bonds



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### BCMB 3100 - Chapter 3

- Polypeptides: definition, structure, and direction
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- Disulfide bonds
- **Protein purification**
- Methods to determine amino acid composition, cleavage of proteins, protein sequencing

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### PROTEIN PURIFICATION

#### General strategy

Tissue → disrupt → crude fractionation → selected fractionation

#### Proteins can be separated by:

- \_\_\_\_\_ : SDS gel electrophoresis, gel filtration chromatography, dialysis, centrifugation
- \_\_\_\_\_ : salting out
- \_\_\_\_\_ : ion-exchange chromatography
- \_\_\_\_\_ : interaction chromatography, reverse phase chromatography
- \_\_\_\_\_ : affinity chromatography, antibodies

#### Purified proteins can be analyzed separated by:

- Sequencing:** Edman degradation, (proteolysis)
- 3D structure:** X-ray crystallography, NMR
- Synthesis:** automated solid phase

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To purify large amounts of proteins one requires:

1. An \_\_\_\_\_ for the protein (enzyme, antibody, etc)
2. A \_\_\_\_\_ to separate desired protein from "all" other proteins and which retains protein "activity"
3. Example strategy:

Salting-out: the "specific" precipitation of a given protein at a specific high-salt concentration

Ion exchange → Gel-filtration → Affinity chromatography

Dialysis often used to remove salts: separation of protein from small molecules through a semipermeable membrane (cellulose)

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### Different types of chromatography (1)

\_\_\_\_\_: proteins passed over a column filled with a hydrated porous beads made of a carbohydrate or polyacrylamide polymer (large molecules exit first)

\_\_\_\_\_: separation of proteins over a column filled with charged polymer beads (+ charge = anion-exchange; - charge = cation exchange. Positively charged proteins bind to beads of negative charge & vice versa. Bound proteins are eluted with salt. Least charged proteins will elute first.

\_\_\_\_\_: proteins are passed through a column of beads containing a covalently bound high affinity group for the protein of interest. Bound protein is eluted by free high affinity group.

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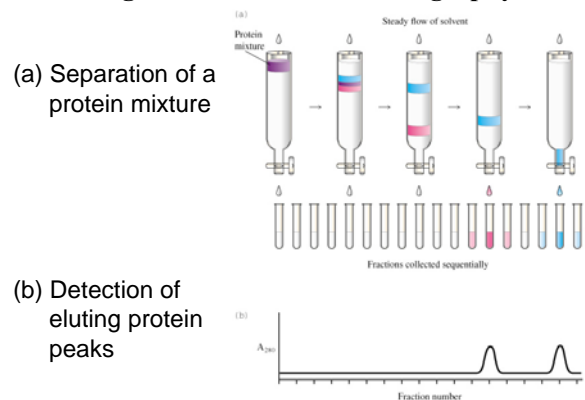
### Different types of chromatography (2)

**Chromatography based on hydrophobicity of protein:**

\_\_\_\_\_ (HIC) and \_\_\_\_\_ (RPC) are both based on interactions between hydrophobic patches on the surface of a protein and on the hydrophobicity of ligands (e.g. alkyl groups) covalently attached to a gel matrix. In RPC proteins can bind very strongly to the gel and require non-polar solvents for their elution. In HIC protein binding is promoted by inclusion of salt in the solvent and elution of proteins is caused by decreasing or removing salt from the solvent.

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Fig 3.11 Column Chromatography

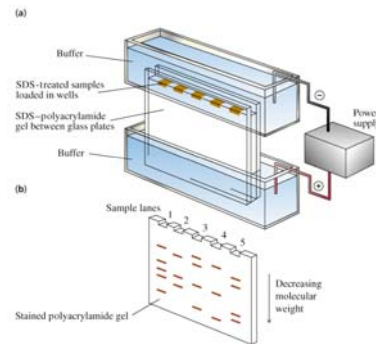


\_\_\_\_\_ : movement of charged solutes through a gel in response to an electric field

\_\_\_\_\_ :  
chemically inert; polymerized acrylamide matrix of controlled pore size; allows separation of proteins based on mass and charge

\_\_\_\_\_ : (sodium dodecyl sulfate, page 30): anionic detergent used for polyacrylamide gel electrophoresis. It complexes with proteins (1 SDS/2 amino acids) → denatured protein of negative charge proportional to protein mass. Note: reducing agents (mercaptoethanol, dithiothreitol) are also added to reduce disulfide bonds. Mobility of many proteins under these conditions is linearly proportional to the log of their mass. Smaller proteins migrate faster.

Fig 3.12 (a) SDS-PAGE Electrophoresis  
(b) Protein banding pattern after run



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Proteins can also be separated by electrophoresis based on their native charge.

\_\_\_\_\_ : pH at which net protein charge is zero

\_\_\_\_\_ : electrophoresis of proteins (w/o SDS) in a pH gradient to a position in the gel at which pH = pI. pH gradient formed by polyampholytes (small multi-charged polymers of many pIs).

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- Polypeptides: definition, structure, and direction
- Peptide bond
- Disulfide bonds
- Protein purification
- Methods to determine amino acid composition, cleavage of proteins, protein sequencing
- Diversity of proteins

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### Amino Acid Composition (1)

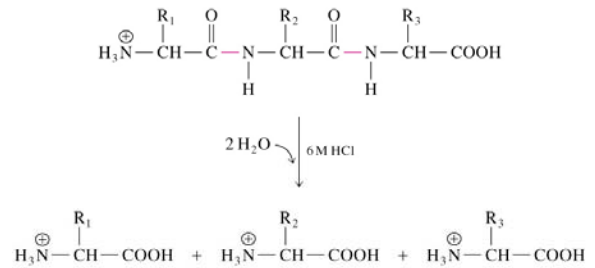
#### 1. Determine amino acid composition

hydrolysis  
**a. peptide** →→ free amino acids  
 6 N HCL  
 100°C, 24 hr

**b.** Amino acid composition of hydrolysates determined by automated cation-exchange chromatography (amino acid analyzer). Column contains solid granules of sulfonated polystyrene. Amino acids reacted with ninhydrin (yields colored product) & detected by O.D.

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**Fig 3.15 Acid-catalyzed hydrolysis of a peptide**



Pg 72

**Problem: Asn → Asp; Asn + Asp = Asx or B  
 Gln → Glu; Gln + Glu = Glx or Z  
 Loose some: Ser, Thr, Tyr; Loose most Trp**

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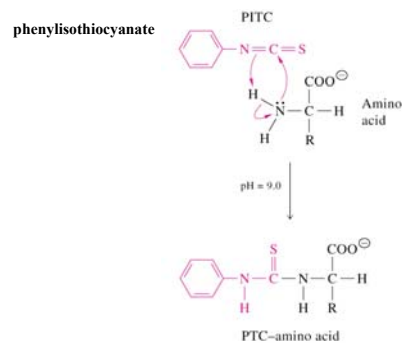
### Amino Acid Composition (2)

**c.** Another method for aa composition analysis is to treat protein hydrolysate with phenylisothiocyanate (PITC) at pH 9.0 to yield PITC-aa derivatives, separate by HPLC via hydrophobic attraction of aa side chains to hydrocarbon matrix of column and quantitate by OD 254nm (due to PTC moiety).

(The first pure protein analyzed for a.a. composition was β-lactoglobulin. It took several years of work. Today amino acid analyzer allows composition analysis within 2-4 hours with samples as small as 1 pmole!!!)

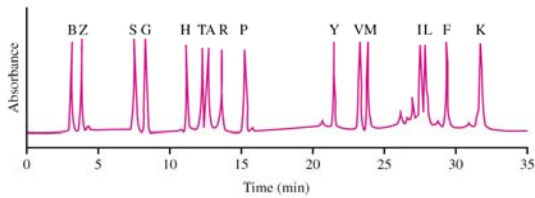
67

**Fig 3.16 Amino acid treated with PITC**



68

**Fig 3.16 Chromatogram from HPLC-separated PTC-amino acids**



Note: acid treatment converts Asn→Asp; Gln→Glu; Trp is destroyed, some loss of Ser, Thr, Tyr.  
 B = (Asn→Asp) + Asp; Z = (Gln→Glu) + Glu

69

## 2. Determine amino-terminal residue

Sanger devised the first method to label N-terminal residue by reacting 2,4-dinitrophenylbenzene with  $\alpha$ -amino group →→ yellow derivative. Subsequent hydrolysis (6N HCl) hydrolyzes away all other amino acids. N-terminal residue derivative identified by chromatography

Today Dabsyl chloride (→colored derivative) or Dansyl chloride (→ fluorescent derivative) are used.

70

## 3. Determination of amino acid sequence by Edman Degradation (Pehr Edman) (1950)

Edman Degradation is now done on sequencers (automated, Liquid phase sequenator →→ analysis by HPLC (one cycle 2 hr)

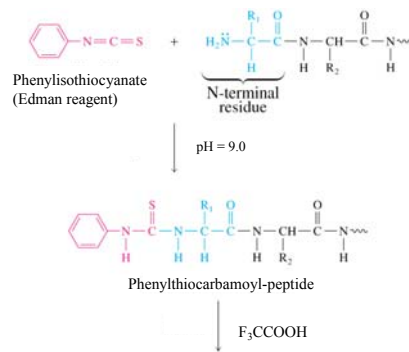
Gas-phase sequenator: detection of pmole amounts (single SDS-PAGE band)

Frederick Sanger determined the first complete sequence of a protein (insulin) in 1953 (51 amino acid long)

Note: Amino acid sequence of small proteins and peptides is now commonly determined by mass spectrometry (e.g. electrospray MS, MALDI MS).

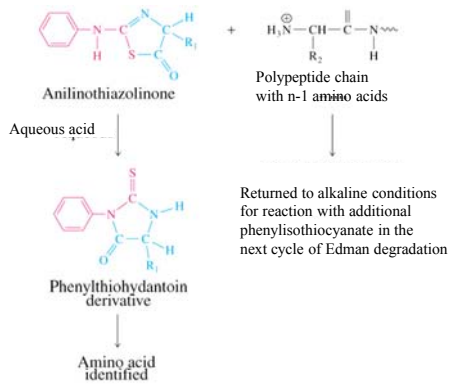
71

**Fig 3.18 Edman degradation procedure**



72

### Edman degradation procedure (cont)



73

### Protein Cleavage

Edman degradation limited to polypeptides of 50 a.a.

Chemical cleavage:

\_\_\_\_\_ splits on carbonyl side of Met

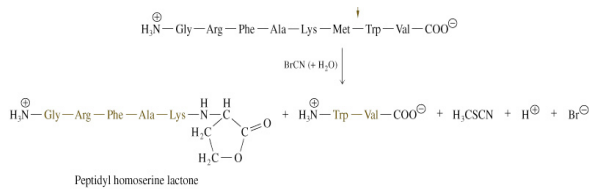
Enzymatic cleavage:

\_\_\_\_\_ : protease cleaves on carbonyl side of Arg and Lys

\_\_\_\_\_ : protease cleaves on carbonyl side of bulky hydrophobic and aromatic amino acids

74

Fig 3.20 Protein cleavage by BrCN



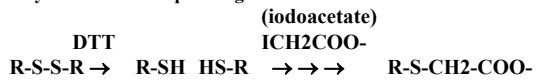
75

Fig 3.21 Cleavage, sequencing an oligopeptide



76

Disulfide bonds must be removed from protein by reduction & alkylation before sequencing.

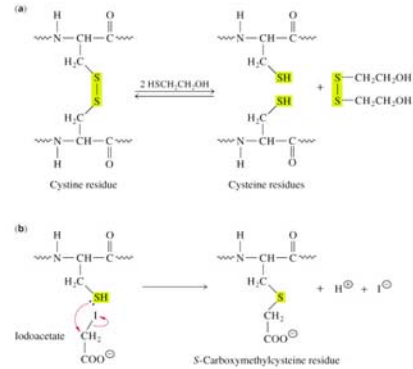


DNA recombinant technology → DNA sequence of nascent protein

\_\_\_\_\_ : direct polypeptide product of translation (no modification)

77

Fig 3.19 Cleaving, blocking disulfide bonds



78

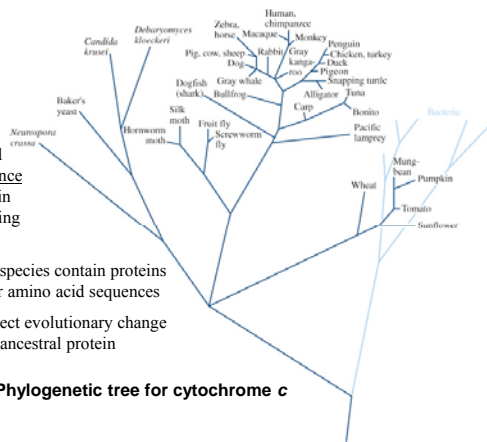
Fig 3.24  
(see also  
Fig. 3.23)

•Protein amino acid sequences can be deduced from the sequence of nucleotides in the corresponding gene

•Closely related species contain proteins with very similar amino acid sequences

•Differences reflect evolutionary change from a common ancestral protein sequence

Phylogenetic tree for cytochrome c



## Polypeptide chain nomenclature

- Amino acid “residues” compose peptide chains
- Peptide chains are numbered from the N (amino) terminus to the C (carboxyl) terminus
- Example: (N) Gly-Arg-Phe-Ala-Lys (C) (or GRFAK)
- Formation of peptide bonds eliminates the ionizable  $\alpha$ -carboxyl and  $\alpha$ -amino groups of the free amino acids

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